



Kinetics Release Notes

Updated August 2025

Kinetics 2.0

Release August 2025

The latest Carterra Kinetics and Epitope Analysis Software brings many exciting feature updates to enhance ease-of-use and improve the user experience when analyzing large data sets.

Please carefully read the following notes before downloading for use. Additional details about the software and platforms can be found in the User Manual, available for download: <https://carterra-bio.com/resource-category/user-manuals/>

Changes Since Last Release

Features

- 2.0 is compatible with files from all Carterra instruments.
- Calibration files can be exported from one analysis and imported into another. Make sure to only do this when chips and buffers have not been changed.
- Crop can now be applied prior to calibration.
- Custom referencing is supported for non-standard formatting of arrays.
- Kinetics and Epitope now support calibration; response normalization over the full range of your data.
- Kinetics supports the Excluded Volume Correction, in support of small-molecule applications.
- NEW: kinetics screening tab. Customizable hit selection.
- Steady State tab now support Y-axis data normalization.

Updates

- "ROI position and ROI ID" nomenclature has been replaced with ROI.
- Analyte filtering before calibration is now possible
- Data sorting now first sorts on ascii, then numeric
- Exclusion/inclusion of data in kinetics tab no longer causes the table to jump back to the beginning. Continue analyzing and curating your data from where you left off after hiding/unhiding analyte/ligand pairs.
- Exported tables are now labeled with units for all columns.
- Exporting to Excel and SQLite has been accelerated.
- Files now open faster than ever, the application closes faster, fitting speed has been improved, and navigation is more fluid and responsive.
- Merge+save now reports the location of the saved file.
- More user control of data display and export options in gears tabs.
- Parameter precision now based on fit confidence.
- Quantitation and surface prep arrays files have their immobilization phases broken up like capture kinetics for consistency.
- Rendering speed has been enhanced.
- Hardware rendering issues introduced by changes in Intel drivers have been resolved.
- ROI map now fits in view.
- Summary stats page now uses geometric mean to average the on- and off-rate data. The average K_D is now reported as the ratio of $k_d(\text{geomean})/k_a(\text{geomean})$, not the average of the individual K_D measurements.
- Version 2.0 no longer supports .isadata files. Files before ~June 2020 will need to be opened with legacy Kinetics or Epitope.
- Parameter error analysis has been improved.

Calibration and Excluded Volume Correction

Note: At this time, the Calibration and EVC features are exclusive to the Catterra Ultra platform.

Contents

This protocol provides guidelines for designing and executing Calibration and Excluded Volume correction in Navigator software.

Calibration

Calibration utilizes responses generated from twelve calibrant solutions spanning a range from low to high refractive index (RI) (relative to assay conditions). This procedure linearizes the ΔRU vs. ΔRI scale for all experimental and reference ROIs across the chip. Calibration should be performed for each experiment where the highest data quality is desired (single point screens, affinity/kinetics, epitope binning). This workflow can be performed as part of a wizard or set up manually by user.

Calibration solutions should be optimized for each experiment. Here are some suggestions:

low RI = 4.5 mL water or buffer-DMSO + 10.5 mL running buffer

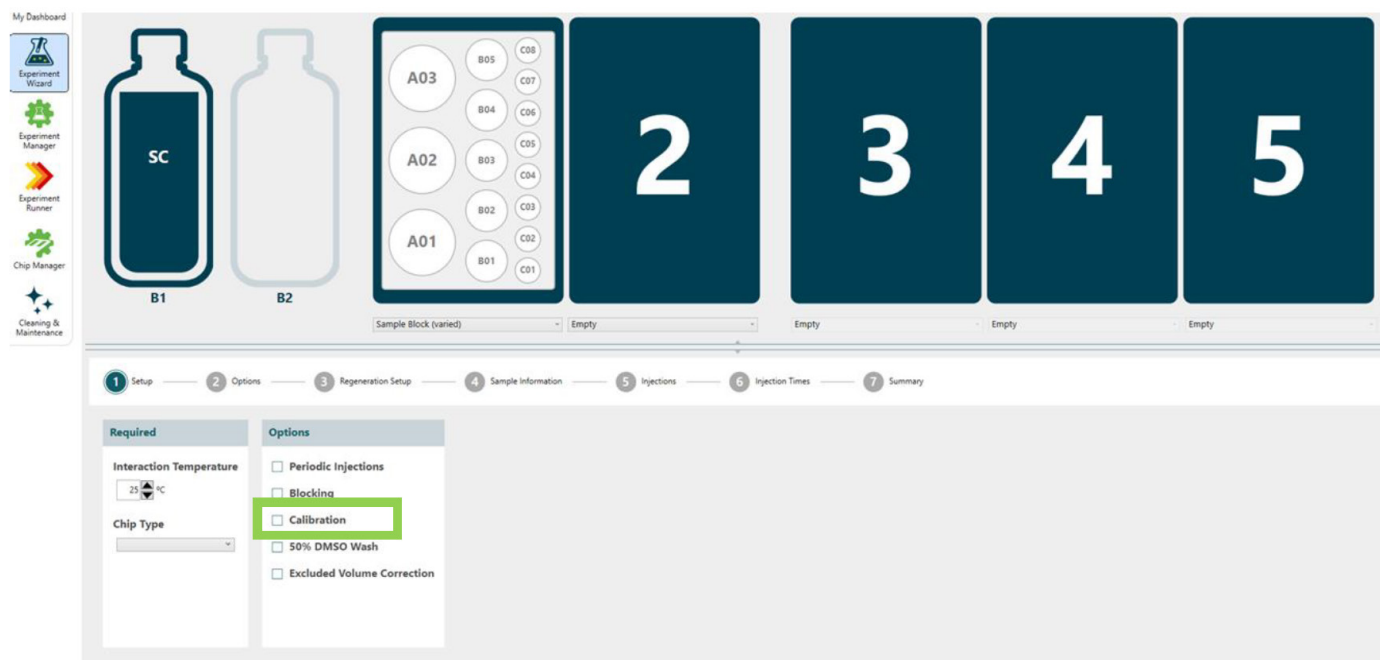
high RI = 600 μ L DMSO + 14.4 mL running buffer

Other solutions can be used in place of DMSO as a high RI component. For example adding 1,200 μ L of 50% glycerol or 3 mL of 5 M NaCl will result in similar responses.

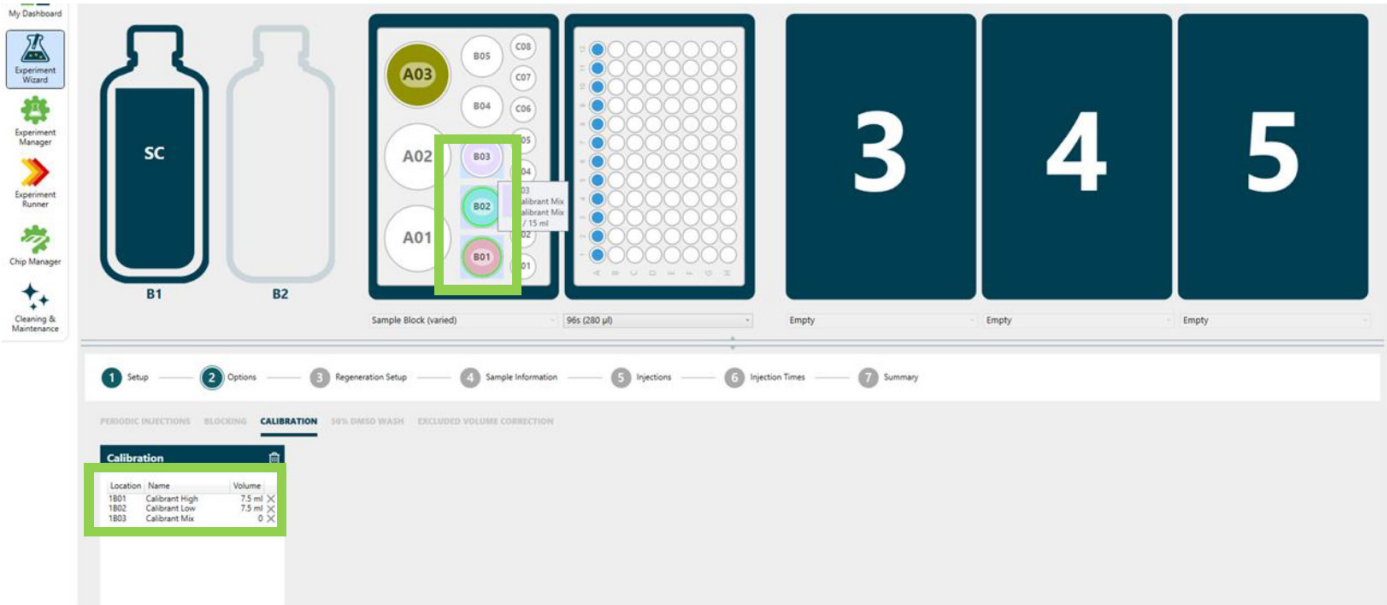
Utilizing Calibration option in Navigator:

Automatic calibration is performed at the end of the experiment and takes approximately 45 minutes. This calibration procedure requires user to supply at least 7.5 mL of well-mixed high and low RI solution and an empty vial for mixing. Ultra will automatically generate and inject twelve mixed samples from the two provided solutions. Below are the steps for setting up automatic calibration:

1. Calibration option can be selected from the Setup tab of Experiment wizard.



2. Proceed to Options tab to populate sample locations. Calibration requires selection of three positions with minimum capacity of 7.5 mL (select from Sample block locations in A or B). The positions will be assigned in the order selected: high RI, low RI, mix. In example below positions used are B1 (high RI), B2 (low RI), and B3 (mix).



3. Prepare at least 7.5 mL of high and low RI solutions to include in B1 and B2, respectively. Place empty 15-mL vial into position B3.

Manual calibration setup:

When using microplates in both Single Channel bays (or when user is unable to designate 3 large volume vials), automatic calibration is not possible. Calibration can still be performed by preparing calibration solutions manually and including them in the injection list. To perform manual calibration, follow these steps:

1. Do not select Calibration option from the Setup tab of Experiment wizard.
2. Prepare a set of twelve solutions by mixing various volumes of the high and low RI solutions as follows:

		Calib 1	Calib 2	Calib 3	Calib 4	Calib 5	Calib 6	Calib 7	Calib 8	Calib 9	Calib 10	Calib 11	Calib 12
Parts	Low RI	11	10	9	8	7	6	5	4	3	2	1	0
	High RI	0	1	2	3	4	5	6	7	8	9	10	11

Or

		Calib 1	Calib 2	Calib 3	Calib 4	Calib 5	Calib 6	Calib 7	Calib 8	Calib 9	Calib 10	Calib 11	Calib 12
Volume (uL)	Low RI	220	200	180	160	140	120	100	80	60	40	20	0
	High RI	0	20	40	60	80	100	120	140	160	180	200	220

3. Include minimum of 200 μL of each solution in the plate.

The screenshot shows the 'Injections' step in the Kinetics software. The top part of the interface displays a schematic of the experimental setup, including two sample containers (B1, B2), two 96-well plates (each containing 200 μL of solution), and three large blue boxes labeled 3, 4, and 5. The bottom part of the interface shows a table of injection data for Bay 1 and Bay 2. The table has columns for Location, Name, Conc (M), MW (Da), Refractivity (I), and Molecular Vt. Notes. The 'Injections' step is highlighted in the top navigation bar.

4. Add calibrant solutions to SC injections list at the end of the sample injections.

The screenshot shows the 'Add Injections' dialog box and the 'SC Injections' table. The 'Add Injections' dialog box has three sections: 'Verify used wells:', 'Sorting:', and 'Preview order:'. The 'Verify used wells:' section lists wells H01 through H12, all of which are checked. The 'Sorting:' section has three options: 'By Row (Letters)', 'By Column (Numbers)', and 'Custom'. The 'Preview order:' section shows a list of wells (2H01 through 2H12). The 'Add 12 Injections' button is highlighted in green. The 'SC Injections' table shows a list of injections (151-164) with columns for Injection #, Bay, Location, Name, and a status icon. An arrow points from the 'Add 12 Injections' button in the dialog box to the 'SC Injections' table.

5. During data processing in Kinetics, highlight the 12 calibration injections, right click, and assign to group "Calibrants."

Excluded Volume Correction (EVC)

Note: At this time, the Calibration and EVC features are exclusive to the Catterra Ultra platform.

EVC accounts for the refractive index shift that is observed between experimental and reference surfaces caused by the captured/coupled material in the experimental region taking up volume in the detection region that is unoccupied on the reference surfaces. More of those buffer component molecules fit in the reference region leading to a refractive index shift that is independent of any binding interactions. This effect is exaggerated by high coupling densities as well as the use of high refractive index solvents such as DMSO, though in principle is present in all experiments. This correction is important for low- and high-affinity interactions but is most critical when no kinetics are visible and the binding interaction and bulk-shift components become indistinguishable from each other.

EVC requires at least 6 solutions (recommended 7-12 solutions) with a minimum volume of 200 μL . Low and high RI solutions can be prepared similarly to what is described in the Calibration section. Ensure that EVC falls within the Calibration RI span. EVC solutions must be prepared manually and included in Injections list. Below is the suggested procedure for preparing 8 EVC solutions using the same low and high RI solutions as for calibration:

		EVC 1	EVC 2	EVC 3	EVC 4	EVC 5	EVC 6	EVC 7	EVC 8
Parts	Low RI	8	7	6	5	4	3	2	1
	High RI	1	2	3	4	5	6	7	8

or

		EVC 1	EVC 2	EVC 3	EVC 4	EVC 5	EVC 6	EVC 7	EVC 8
Volume (μL)	Low RI	800	700	600	500	400	300	200	100
	High RI	100	200	300	400	500	600	700	800

*Note the EVC 1 and 8 are mixes of low and high RI solutions, which allows them to stay within the Calibration range.

Below are the steps for setting up EVC:

1. Select EVC option in the Setup tab of Experiment wizard.

My Dashboard

Experiment Wizard

Experiment Manager

Experiment Runner

Chip Manager

Cleaning & Maintenance

SC

B1

B2

A03

A02

A01

B05

B04

B03

B02

B01

C08

C07

C06

C05

C04

C03

C02

C01

Sample Block (varied)

Empty

Empty

Empty

Empty

1 Setup

2 Options

3 Regeneration Setup

4 Sample Information

5 Injections

6 Injection Times

7 Summary

Required

Options

Interaction Temperature

25 $^{\circ}\text{C}$

Chip Type

CMDP

☐ Periodic Injections

☐ Blocking

☒ Calibration

☐ 50% DMSO Wash

☒ Excluded Volume Correction

SC

B1

B2

A03

A02

A01

B05

B04

B03

B02

B01

C08

C07

C06

C05

C04

C03

C02

C01

Sample Block (varied)

96s (200 μL)

Empty

Empty

Empty

1 Setup

2 Options

3 Regeneration Setup

4 Sample Information

5 Injections

6 Injection Times

7 Summary

PERIODIC INJECTIONS

BLOCKING

CALIBRATION

50% DMSO WASH

EXCLUDED VOLUME CORRECTION

Excluded Volume Correction

Location	Name	Volume
2H01	Excluded Volume Corre	0 \times
2H02	Excluded Volume Corre	0 \times
2H03	Excluded Volume Corre	0 \times
2H04	Excluded Volume Corre	0 \times
2H05	Excluded Volume Corre	0 \times
2H06	Excluded Volume Corre	0 \times
2H07	Excluded Volume Corre	0 \times
2H08	Excluded Volume Corre	0 \times

2. Under Options tab in Excluded Volume Correction subtab, add a minimum of 6 EVC positions to Bay 1 or 2.
3. On Injections tab, add EVC solutions to the end of the injections list.

The screenshot displays the Fluidigm C1 software interface during the 'Injections' step. The main workspace shows a chip layout with two blocks, B1 and B2. Block B1 contains a sample labeled 'SC'. Block B2 is empty. A sample block is shown with a varied sample, and the volume is set to 96s (200 µl). Three empty injection wells are visible, labeled 3, 4, and 5. A progress bar at the bottom indicates the current step is 5, 'Injections'. A table titled 'SC Injections' is shown, listing injection details for various samples. A green box highlights rows 37-44, which are marked as 'Excluded Vo'.

Injection #	Bay	Location	Name
31	2	C07	2C07
32	2	C08	2C08
33	2	C09	2C09
34	2	C10	2C10
35	2	C11	2C11
37	2	H01	Excluded Vo
38	2	H02	Excluded Vo
39	2	H03	Excluded Vo
40	2	H04	Excluded Vo
41	2	H05	Excluded Vo
42	2	H06	Excluded Vo
43	2	H07	Excluded Vo
44	2	H08	Excluded Vo

Kinetics 1.9.2.4463

Release May 2024

- Features
- Updates
- Known Issues

Changes Since Last Release

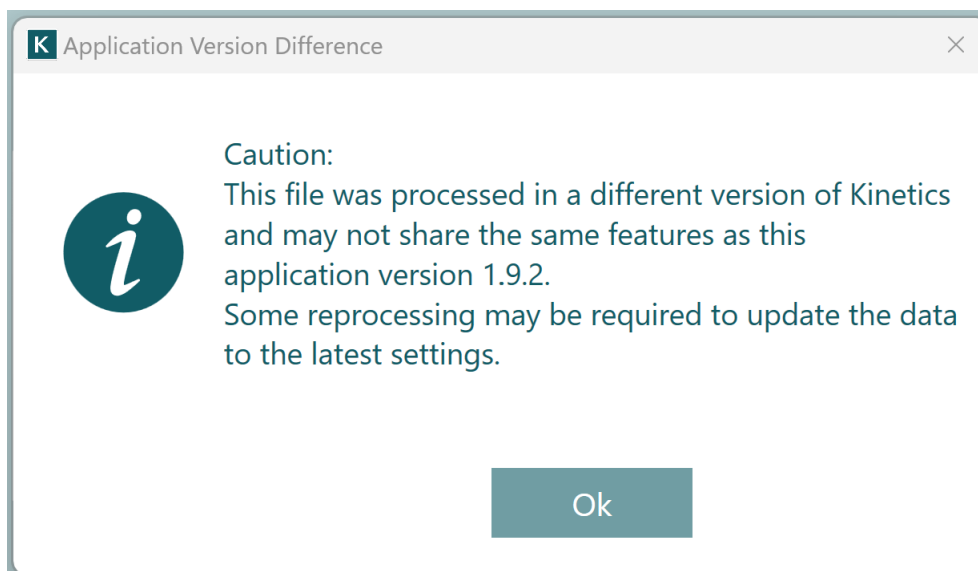
Features

• Misc

- Updates made to application installer to recognize existing installed Microsoft C++ redistributables and simplify installation dialog.
- Kinetics 1.9.2.4463 will be the last version to support the .lsadata file format.

• File Opening and Saving

- If files saved in previous versions are opened, caution message will appear about recommended best practices when working with the file in this latest version.



• Data Tab

- Reassignment of Analytes in right-click menu is restricted based on original assignment (Analyte or Immob) to avoid errors.

• Processing Tabs

- ROI map view now uses experimental coordinates indicating the actual placement of ROIs on the chip during the run. Note that for previously processed files the ROI map will revert to a default layout.
- ROI map is now draggable and zoom in/zoom out can be done by CTRL+ mouse wheel.

▪ Data Fitting

- Fitting progress dialog now included when fitting data on the Kinetics>Kinetic Analysis tab.

Fitting progress

Current executing tasks:

- Fitting
- Visualizing

Cancel

- Default k_a fitting uses range of starting values to avoid fitting minima which impact accuracy of k_a fits.
- Users have option of instead checking k_a box on Kinetics tab and assigning a specific k_a starting value.

- Show K_D Units is now unchecked by default.

▪ Analysis Tab

- No new features.

▪ Report Point Tab

- No new features.

▪ Data Export

- Headers are no longer merged in the Excel export.
- Analyte and Ligand Report Point tables are now separate tabs in Excel export.
- With Show K_D Units now unchecked by default, no units are included in the export tables.

Updates

▪ Misc

- No updates.

▪ File Opening and Saving

- No updates.

▪ Data Tab

- Duplicate prints now correctly separated into discrete Link Groups.
- Correction of bug allowing space to be appended at end of Ligand names.
- Carryover of ligand names from previous experiments in block 1 resolved.

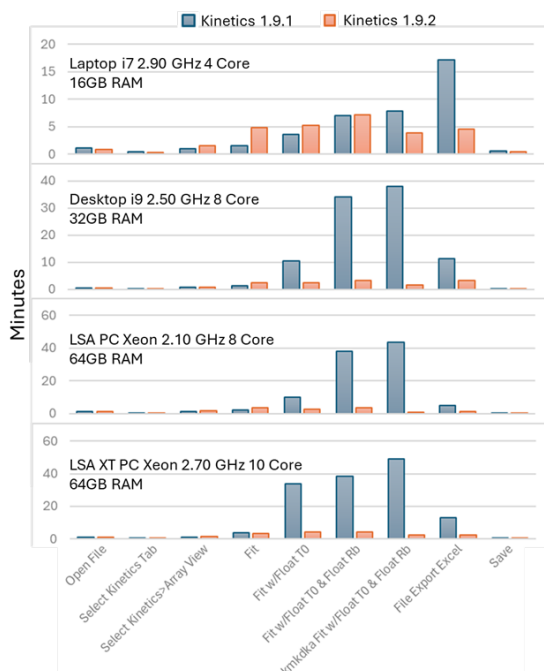
▪ Processing Tabs

- Corrections made to custom and global referencing options to ensure displayed referencing assignments are applied appropriately.

▪ Data Fitting

- T0 is now functional when analyzing previously analyzed files without requiring deselecting, fitting, then reselecting. Carterra still recommends refitting processed files when opened in the latest builds to ensure most recent improvements are applied to the data.

- Fitting speed performance improvements implemented which reduce fitting steps by up to an order of magnitude. See chart below for examples of performance when fitting a 16-link group file on various PC types in Kinetics 1.9.1 vs. 1.9.2.



- Flagging of data in the Steady State tab only highlights and no longer removes data from the view.

• Analysis Tab

- Issues with data not fully displaying on Isoaffinity chart have been resolved.
- Errors when sorting on the Affinity tab have been resolved.

• Report Tab

- No updates.

Known Issues

- Report point export tables for Immobilization are missing Bay information .
- Data flagged as < 50% Rmax, kd below limit, and/or km below limit is not treated as Flagged in the Analysis tabs.
- Characters or numbers other than numbers 1 through 432 added to the Global Reference in the Analyte and Ligand Processing tabs will cause an error to appear.
- When Normalization is applied in Steady State, chart autoscaling may not apply correctly and therefore scales should manually defined.
- However, in previously processed files, the ROI map will open with a default layout that is not consistent with the physical layout on the chip. Referencing relationships are correct.
- When running Capture Kinetics without specifying prints, the data file will open with default names of Ligand 1, Ligand 2, etc., and the ligand names will need to be manually added to the file.

Kinetics 1.9.1.4215

Release October 2023

Changes Since Last Release

- Features.
- Updates.
- Known Issues.

Features

- Allow k_d fitting for multichannel (immob) data.
- Data displayed during multichannel rinsing phase.
- Chart export now accounts for screen scaling.
- Ability to correct sensorgram bulk shift.
- Display data within analyte injection cycle.
- Kinetic fitting routine code improvements.
- Kinetics and Steady State pages, Array View tabs: make "Show ROI Position" and "Show Ligand Name" clicked on by default. Update Settings > Table Options > Display ROI to be Checked by Default.
- Gear menu options to hide Ligand Name and ROI from Array View.
- Add Global Referencing to Analyte Processing Page.
- K_D header row moved in File>Excel Export to simplify table processing.

Updates

- Link Groups not correctly delineating.
- Tiles in File>Export do not follow sorted order in application.
- High residuals in some off-rate fits.
- An unstable error appears when opening certain saved .kitx files.
- Incorrect sorting option by ROI ID in the Kinetic Analysis, Global Analysis tables.
- Analysis > Stats page: Axis data is clipped.
- Incorrect behavior of "ka" gates on the Analysis > Iso-Affinity page.
- The X and Y axis label size settings don't apply to the chart on the Analysis > Stats page.
- The chart text size settings don't apply to the chart on the Analysis > Scatter Plot page.
- The Inactive ligands are displayed in the "Analytes" tab on the Analysis > Stats page.
- Application isn't updated dynamically after using the "Data Filter" option.
- Experiments are not displayed on the charts on the Analysis pages when the experiment assigning to the Default group from the Inactive group on the Steady State page.
- Sort order changed after experiments are assigned to the Inactive / Complex / Excluded Group in the Kinetic Analysis table.
- "Copy All Data to Clipboard" Error on Kinetics > Array View.
- Scatter plot pasted values don't match plot.
- Data page: Font sizes do not change.
- SQLite Export missing ResSd value from fitting results.
- Cannot Copy/Paste into Analyte Processing > Referencing > Assignments Table.
- Steady State > Global Analysis Table Missing "ROI" Column.
- Quant Page - Report "[Concentration Value] <,> [Max/Min Allowable Concentration]" instead of ND.
- After fitting on the steady state page some ligands with ND values are not colored as Inactive.

Known Issues

- Fitting times increase as more fit variables/options are selected.

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